**Title**: Plant symbiotic fungal isolate of *Alternaria alternata* grows better with an inorganic nitrogen source than an organic nitrogen source

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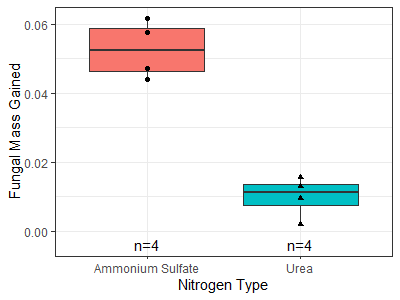
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**Abstract**

<100 words

*Alternaria alternata* is a fungus that causes diseases of agriculturally important plants, but it is also a Dark Septate Endophyte (DSE), a group of fungi that can improve the growth of host plants often by improving access to soil resources like nitrogen. To facilitate experimental testing of the environmental factors that influence this relationship, it is necessary to know if *A. alternata* can use both organic and inorganic nitrogen. We found that an *A. alternata* isolate grew 81% larger in an inorganic nitrogen medium than in an organic nitrogen medium. These data provide grounds for future studies testing more DSE taxa and more forms of organic and inorganic nitrogen to assess how different fungal groups utilize nitrogen.

**Figure**



**Figure caption**: Fungal biomass growth was significantly higher in media containing an inorganic nitrogen source compared to an organic nitrogen source (Welch Two Sample t-test; meanAmmonium Sulfate = 0.0525975; meanUrea = 0.0099475; t(5.3753) = 8.3487, p = 0.0003). Points are shaped according to nitrogen type and indicate each replicate? in the experiment.

**Description**

*Alternaria alternata* is a fungus that causes a spot disease in many plant species , a concern because of the substantial declines in crop productivity that it can cause (Troncoso-Rojas & Tiznado-Hernández, 2014). *A. alternata* alsois considered a Dark Septate Endophyte (DSE), which is a polyphyletic group of ascomycete fungi defined by regular cross-walls among cells (septa) and darkly pigmented hyphae (DeMers, 2022). While many DSE have been categorized as plant pathogens, there is increasing evidence that under certain conditions, these fungi can form mutualisms with plants and improve plant growth through improved access to soil resources including nitrogen, instead of causing disease (Berthelot et al., 2019; Netherway et al., 2024; Schulz & Boyle, 2005). Given that *A. alternata* is a globally distributed species (DeMers, 2022), it is of interest to understand the conditions that are conducive to its function as a mutualist instead of a pathogen so its management in agricultural settings can be improved.

In this experiment, we assess the ability for an *A. alternata* isolate (GenBank Accession Number SUB14593255 germinator\_fung\_10x\_B\_2\_F12 PQ284877) to utilize inorganic and organic forms of nitrogen. It is vital to know if this fungus can use both forms of nitrogen because this will impact which forms of nitrogen are used and monitored in experiments. For example, stable isotopes of nitrogen are useful in tracing nitrogen movement in plant-fungal symbioses. However, it is unclear which form of nitrogen is best to use for these experiments despite previous work on other groups of fungi (Finlay et al., 1992; Hawkins et al., 2000; He et al., 2003; Newsham, 2011; Upson et al., 2009). Additionally, nitrogen is an important nutrient for plant growth, and its use by different fungi and plants is important in agricultural settings where resources and fungal growth are monitored closely.

In this laboratory experiment, we grew the *A. alternata* isolate in nitrogen-free media and amended it with either organic or inorganic nitrogen. We hypothesized that the fungus would be better able to use organic forms of nitrogen because of the fungus being a plant-symbiont which improves plant access to nitrogen, although through unclear mechanisms (Di Martino et al., 2022; Newsham, 2011). In contrast to our expectation, the fungal isolate’s growth was significantly higher in the inorganic nitrogen media than in the organic nitrogen media (Welch Two Sample t-test; meanAmmonium Sulfate = 0.0525975; meanUrea = 0.0099475; t(5.3753) = 8.3487, p = 0.0003). While other sources of inorganic and organic nitrogen should be tested to determine fungal preference between the two types of nitrogen, it is clear that the fungus not only utilized an inorganic nitrogen source, but it also grew better with the inorganic nitrogen source than with the organic nitrogen source in this experiment.

This finding is relevant to a multitude of experiments, including those which aim to use stable isotopes of nitrogen to trace the movement of nitrogen, as the fungus’s ability to use the nitrogen source is vital in such a method. Additionally, while more DSE isolates should be tested for their abilities to use inorganic and organic nitrogen, this experiment provides foundational evidence for DSE being able to use inorganic nitrogen sources.

**Methods**

We prepared two nitrogen-free media solutions by dissolving 0.0274 grams of Nitrogen free minimal media (MyBioSource Edinburgh) per 1 mL of autoclaved RO water. These solutions were amended with either an organic or an inorganic nitrogen source. For the inorganic N treatment, we added 0.0019 grams of Ammonium Sulfate (Mallinckrodt Chemical Works) per 1 mL of solution. For the inorganic N treatment, we added 0.0009 grams of Urea (Fisher Scientific U-15) per 1 mL of solution. Ratios were calculated based on the number of N atoms by weight so as to ensure the same amount of nitrogen was in each treatment. Both solutions were autoclaved and brought to room temperature over 24 hours.

50 mL Erlenmeyer cell culture flasks with metal caps were autoclaved, and 15 mL of either media type was added to each flask with sterile serological pipettes in a sterile biosafety cabinet using aseptic technique. A XXX mm diameter cork borer was used to punch same-sized inoculum plugs from the growing edge of the fungal culture (GenBank Accession Number SUB14593255 germinator\_fung\_10x\_B\_2\_F12 PQ284877), plated two weeks prior on Potato Dextrose Agar. The dry weight of these plugs is approximately XXX. One fungal plug was added to each flask. Flasks were incubated at 19 ºC shaking at 80 RPM for 7 days. Filter papers were labeled and their individual weights recorded. After 7 days of incubation, the contents of each flask were poured through the filter papers. Flasks were rinsed with autoclaved RO water, and the contents were poured through the same filter papers until the visible contents of all flasks were cleared. Once filtering was complete, we placed folded filter papers upright in a rack in a 59.2 ºC drying oven for 48 hours. After drying, we weighed each sample. Fungal mass was calculated by subtracting each filter paper weight and the average dry fungal plug weight from the final mass of each filtrate.

**Reagents**

*Alternaria alternata* fungus (GenBank Accession Number SUB14593255 germinator\_fung\_10x\_B\_2\_F12 PQ284877). Isolated from a *Sorghum bicolor* seed by Dr. Ron Deckert in Dr. Catherine Gehring’s laboratory at Northern Arizona University.

MyBioSource Edinburgh minimal media, Nitrogen free Catalog #MBS652833

Mallinckrodt Chemical Works Ammonium Sulfate

Fisher Scientific Urea U-15

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Statistical analyses

Fungal mass measurements between nitrogen treatments were compared with a Welch Two Sample t-test. We conducted all data analyses in R version 4.3.0 with RStudio (R Core Team, 2023).

Data availability:

All code and data are available freely at <https://github.com/beabock/Org_vs_Inorg_N>.